

SAMPLING MEDIA

- Any standard-sized (15 x 100mm) Petri dish may be used with the ems E6® Sampler.
- For Bacteria: a Tryptic Soy Agar (TSA) and Tryptic Soy Agar with 5% Sheep Blood or "Blood Agar" (BAP).
- For Fungi: Potato Dextrose Agar (PDA), Malt Extract Agar (MEA), and Dichloran Glycerol 18 Agar (DG-18).

SAMPLE SUBMISSION

- A complete Chain of Custody form should accompany every sample to a lab.
- Samples should arrive in the lab within 24 hours of collection so it is imperative that the sample stay viable; shipping the sample with blue ice or other cooling methods is necessary.



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Bioaerosol Sampler
catalog #120101



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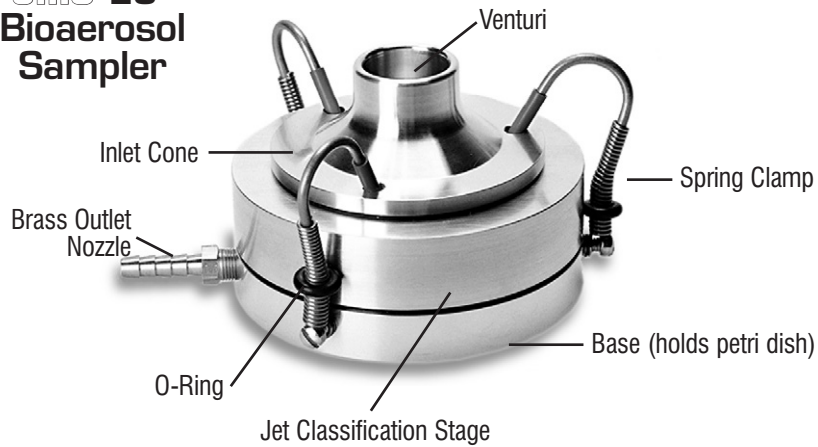
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OPERATING INSTRUCTIONS

ems E6[®] Bioaerosol Sampler



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DESCRIPTION

The ems E6[®] Sampler is a single stage Bioaerosol impactor meeting NIOSH methods 0800 and 0801 for the viable sampling of mold and fungi. The ems E6[®] Sampler is constructed of a corrosion resistant aluminum cone, sampling stage, and base plate secured with aluminum spring clamps and sealed with rubber O-rings. With 400 x 0.25mm precision orifices and state-of-the-art machining techniques, the ems E6[®] Sampler sets the new standard for quality sampling devices.

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REQUIRED EQUIPMENT

- ems E6[®] Bioaerosol Sampler
- Vacuum Pump capable of pulling 28.3 LPM
- Calibration device
- Flexible tubing
- Sampling Media
- Isopropyl Alcohol or other appropriate sampler cleaner
- Hand Sanitizer
- Sterile gauze pads

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SAMPLING METHODOLOGY

1. Hands should be cleaned with an anti-microbial agent prior to setting up, sampling and at any time where cross-contamination may have occurred.
2. Connect one end of the flexible tubing to the vacuum pump and the other end to the ems E6[®] Sampler via the brass outlet nozzle.
3. Calibrate the vacuum pump to 28.3 LPM using a calibration instrument.
4. Use a sterile gauze pad after calibration to wipe all surfaces with isopropyl alcohol or other appropriate sampler cleaner.
5. With the ems E6[®] Sampler sanitized, remove the inlet cone and jet classification stage and place an opened agar plate (with appropriate media) into the base of the sampler and immediately replace the stages and secure with the three spring clamps making sure there is a good seal between the base and the jet classification stage.
6. Turn the vacuum pump on for two (2) to five (5) minutes (depending on type of sample to be collected, call EMS for help if needed). Air will be drawn through the inlet cone into the jet classification stage where it is accelerated through 400 x .25mm holes and impacted onto the agar plate underneath. The exhausted air is then passed through the brass outlet nozzle on the base into the flexible tubing to the vacuum pump.
7. At the conclusion of the sampling, unhook the spring clamps and remove the agar plate. Place the lid on the agar plate immediately and label the plate on the bottom with the appropriate identification. Seal the plate in a zip-lock bag and keep cool.
8. Before taking any other samples, be sure that your hands and sampling equipment have been sanitized to prevent cross-contamination.

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QUALITY CONTROL

- Never use sampling media that has been contaminated, cracked, or is expired.
- Outdoor and indoor "non-affected" area samples should also be collected for comparison to indoor samples.
- Calibrate your vacuum pump as often as necessary to ensure consistent sampling.